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Product Sheet

H_CCR8 Jurkat Cell Line

Catalog number: GM-C15568

Version 3.3.1.241127

Description	H_CCR8 Jurkat Cell Line is a clonal stable Jurkat cell line that constitutively expresses the human CCR8 gene, constructed using lentiviral technology.	
Quantity	5E6 Cells per vial,1 mL	
Product Format	1 vial of frozen cells	
Shipping	Shipped on dry ice	
Storage Conditions	Liquid nitrogen immediately upon receipt	
Target	Human_CCR8	
Gene ID/Uniprot ID	NP_005192.1	
Host Cell	Jurkat	
Recovery Medium	RPMI 1640+10% FBS+1% P.S	
Growth medium	RPMI 1640+10% FBS+1% P.S+0.75 µg/mL Puromycin	
Note	None	
Freezing Medium	90% FBS+10% DMSO	
Growth properties	Suspension	
Growth Conditions	37°C, 5% CO ₂	
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.	
Safety considerations	Biosafety Level 2	
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.	



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Materials

Reagent	Manufacturer/Catalogue No.
RPMI 1640	VivaCell/C3010-0500
Fetal Bovine Serum	Cegrogen biotech/A0500-3010
Pen/Strep	Thermo/15140-122
Puromycin	Genomeditech/GM-040401
Anti H_CCR8 hIgG Antibody	In house/

Figures

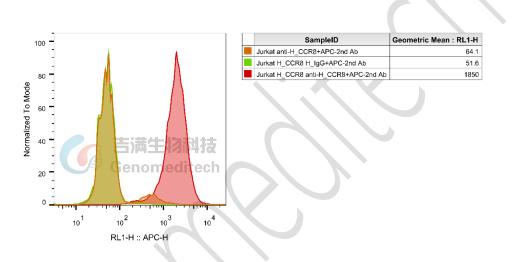


Figure 1 | H_CCR8 Jurkat Cell Line (Cat. GM-C15568) was determined by flow cytometry using Anti-H_CCR8 hIgG Antibody (In house).

Cell Recovery

Recovery Medium: RPMI 1640+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70° C. Storage at -70° C will result in loss of viability.

- a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 3 minutes).
- b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium. And spin at approximately 176 x g for 5 minutes. Discard supernatant.

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- d) Resuspend cell pellet with the recommended complete medium. And dispense the suspension into 1 2 T-25 culture flasks.
- e) Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- a) Centrifuge at 176 x g for 3 minutes to collect cells.
- b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
- c) Aliquot 1 mL into each vial.
- d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: RPMI 1640+10% FBS+1% P.S+0.75 µg/mL Puromycin

Approximately 48-72 hours after the initial thawing, the cells can be passaged for the first time. After this initial passage, the culture medium can be adjusted to growth medium supplemented with antibiotics. If cells are not passaged within 48 hours, it is recommended to add some fresh recovery medium and place the flask horizontally.

- a) When the cell density reaches 1.5 2E6 cells/mL, subculture the cells. Do not allow the cell density to exceed 2E6 cells/mL.
- b) It is recommended to use T-25 flasks for subculturing.
- c) These cells are suspension cells, and it is recommended to use the "half-medium change" method to maintain optimal cell conditions during passaging.
- d) During passaging, you can directly add fresh growth medium to the culture flask, gently pipette to resuspend the cells, and then transfer the cell suspension to a new T-25 flask for continued culture.

Subcultivation Ratio: Maintain cultures at a cell concentraion between 3E5 and 1E6 viable cells/mL.

Medium Renewal: Every 2 to 3 days

Notes

- a) These cells are sensitive to density, so please ensure that the cell density is maintained within an appropriate range during culture and subculturing.
- b) During the first passage, pay attention to the nutrient supply; if not subculturing, make sure to add fresh recovery medium every other day as needed.

Sequence

CCR8 NP_005192.1

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MDYTLDLSVTTVTDYYYPDIFSSPCDAELIQTNGKLLLAVFYCLLFVFSLLGNSLVILVLVVCKKLRSITDVYL LNLALSDLLFVFSFPFQTYYLLDQWVFGTVMCKVVSGFYYIGFYSSMFFITLMSVDRYLAVVHAVYALKVRT IRMGTTLCLAVWLTAIMATIPLLVFYQVASEDGVLQCYSFYNQQTLKWKIFTNFKMNILGLLIPFTIFMFCYIK ILHQLKRCQNHNKTKAIRLVLIVVIASLLFWVPFNVVLFLTSLHSMHILDGCSISQQLTYATHVTEIISFTHCCV NPVIYAFVGEKFKKHLSEIFQKSCSQIFNYLGRQMPRESCEKSSSCQQHSSRSSSVDYIL

Related Products

CCL1:CCR8		
Tango-H_CCR8-CHO-K1 Cell Line	Cynomolgus_CCR8 CHO-K1 Cell Line	
H_CCR8 CHO-K1 Cell Line	H_CCR8 HEK-293 Cell Line	
H_CCR8 U2OS Cell Line	Mouse_CCR8 CHO-K1 Cell Line	
Rhesus_CCR8-eGFP CHO-K1 Cell Line		
Anti-Cynomolgus_CCR8 hIgG1 Antibody (TPP-21360)	Anti-H_CCR8 hIgG1 Antibody(Defucosylated,BMS-986340)	
Anti-H_CCR8 hIgG1 Reference Antibody(BAY-3375968)	Anti-Mouse_CCR8 mIgG2a Antibody	
Anti-H_CCR8 mIgG1 Antibody(GS-1811)	Anti-H_CCR8 mIgG2a Reference Antibody (433H)	
Human CCR8-N1-35 Protein; hFc Tag	Human CCR8-N1-35 Protein; mFc Tag	

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